

S-4-METHYLPHENYL-0,0-BIS[1-BENZOTRIAZOLYL]PHOSPHOROTHIOATE: A VERSATILE PHOSPHORYLATING AGENT

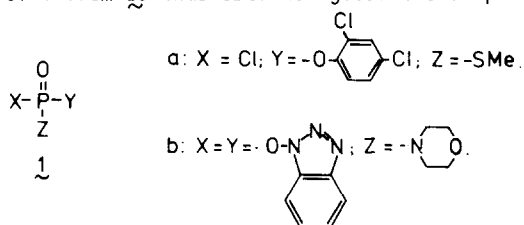
C.T.J. Wreesmann, A. Fidler, G.H. Veeneman, G.A. van der Marel and J.H. van Boom*

Garlaeus Laboratories, PO-BOX 9502, 2300 RA Leiden, The Netherlands

Abstract: The phosphorylating agent obtained by treatment of S-4-methylphenyl phosphorodichloridothioate with 1-hydroxybenzotriazole can not only be applied for the introduction of polyphosphate functions at the terminal ends of nucleic acids, but also for the formation of 3'-5'-phosphotriester linkages.

Nucleic acids which carry terminal polyphosphate groups are from a biological point of view important naturally occurring compounds¹.

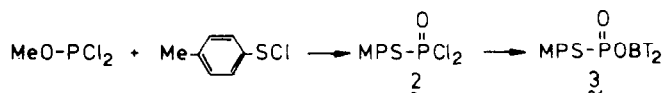
Up to now, two types of phosphorylating agents (i.e. the monofunctional and the bifunctional agents 1a² and 1b³, respectively) have been applied successfully for the introduction of di- and triphosphates at the 5'-end of nucleic acids by a phosphotriester approach. For instance, phosphorylation⁴ of the 5'-OH of a partially protected nucleic acid phosphotriester intermediate (ROH) with 1a affords a 5'-phosphotriester derivative 1a (X=OR; Y=2,4-Cl₂C₆H₃; Z=SMe). Selective removal of Y from 1a thus obtained gives the 5'-phosphorothioate 1a (X=OR;



Y=OH; Z=SMe), which can be converted, according to Nussbaum et al.⁵, into the corresponding 5'-mono, di- or triphosphate derivatives. Recently, we reported⁶ that the phosphorylating agent 1b could serve the same purpose. In this particular case, phosphorylation of a similar 5'-OH function gives the phosphotriester 1b [X=OR; Y=OBT; Z=-N(C₂H₄)₂O] from which the benzotriazolyl (OBT) group can be removed selectively to afford the phosphoromorpholidate 1b [X=OR; Y=OH; Z=-N(C₂H₄)₂O]. The latter can now be converted, according to the original procedure of Khorana et al.⁷, into the required 5'-phosphorylated nucleic acids derivatives.

We now report that S-4-methylphenyl-0,0-bis[1-benzotriazolyl]phosphorothioate (3) is in several aspects more convenient than agents 1a,b and, further, that it can also be used for the formation of 3'-5'-internucleotidic linkages.

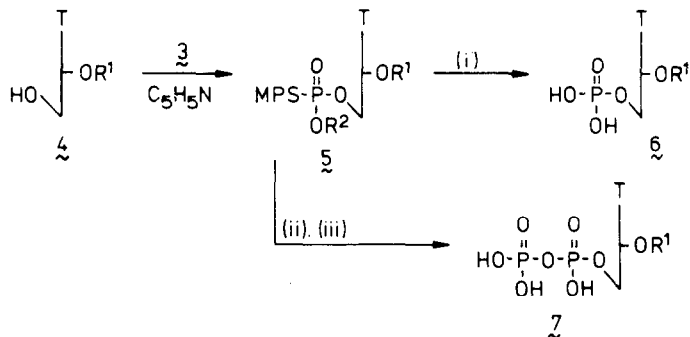
The phosphorylating agent 3 is readily attainable as follows. Treatment⁸ of methoxydichlorophosphine with 4-methylphenylsulfenyl chloride gives, after distillation, 2 as a colourless



solid [Yield: 84%; b.p. 125°C/2 mm Hg; ³¹P-NMR (CDCl₃) δ 33.36 p.p.m.]. A solution of the

latter in dioxan was converted into **3** by the addition of equimolar amounts of pyridine and 1-hydroxybenzotriazole. After 1 h at 20°C, the pyridinium HCl salt was filtered off to give a stock solution (0.2 M) of **3** in dioxan.

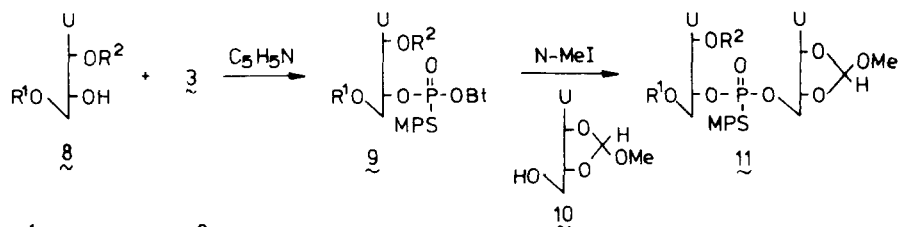
The synthesis of the 5'-phosphates **6** and **7** of thymidine was realized as follows. A solu-



(i): H₂O/I₂. (ii): H₂O/C₅H₅N. (iii): H₃PO₄/I₂.

tion of 3'-O-diphenylacetyl-thymidine⁹ **4** (1 mmol), which was previously dried by coevaporation with pyridine, in dioxan was added to a stirred solution of **3** (1.1 mmol). TLC-analysis, after 15 min at 20°C, showed the absence of **4**. Intermediate **5** (R¹=DPA; R²=BT) thus obtained was hydrolyzed with water-pyridine. Monitoring of this step by ³¹P-NMR spectroscopy revealed instantaneous conversion of **5** (R¹=DPA; R²=BT; δ_p 32.08 and 31.67) into **5** (R¹=DPA; R²=H; δ_p 15.19). Oxidative removal of the MPS-group from **5** (R²=H) to afford the 5'-phosphate **6** (R¹=DPA) was effected by the addition of iodine (10 equiv.) to the reaction mixture. Also in this case, ³¹P-NMR spectroscopy showed a rapid (within 5 min) conversion of **5** (R¹=DPA) into **6** (R¹=DPA; δ_p -0.56). Crude **6** thus obtained gave, after basic hydrolysis of the DPA-group, and consecutive purification by anion-exchange chromatography (DEAE-Sephadex A25) pure **6** (R¹=H; δ_p 0.24) in 80% overall yield. For the preparation of **7**, intermediate **5** (1 mmol; R¹=DPA; R²=BT) was firstly hydrolyzed with water-pyridine and, after 5 min, concentrated to a small volume. The mixture was then repeatedly coevaporated with dry pyridine containing tri-n-butylammonium phosphate (15 equiv.). The formation of the pyrophosphate function was effected by adding iodine (10 equiv.) to the above solution of **5** (R¹=DPA; R²=H). ³¹P-NMR analysis of the reaction mixture showed rapid (5 min) formation of **7** (R¹=DPA; δ_p 8.70, 10.58, J 23.2 Hz). Further processing of crude **7** (R¹=DPA), as described for the preparation of **6** (R¹=H), gave homogeneous **7** (R¹=H) in 75% overall yield.

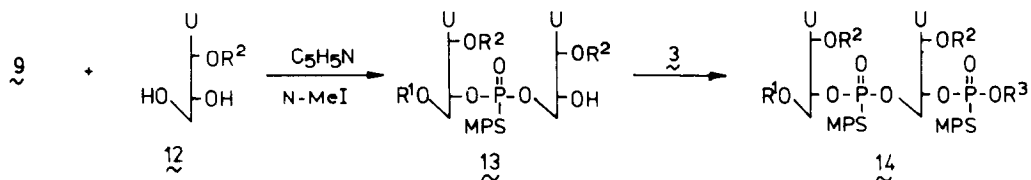
The introduction of a 3'-5'-phosphotriester linkage using **3** was firstly exemplified by the synthesis of the fully-protected RNA dimer **11**. The 2',5'-protected uridine derivative **8**¹⁰



R¹=(C₁₆)DMTr. R²=THP.

(1 mmol) in dry pyridine was treated with $\underline{3}$ (1.15 mmol). TLC-analysis, after 20 min at 20°C, showed complete conversion of $\underline{8}$ into $\underline{9}$ having zero mobility. A solution of 2',3'-protected uridine $\underline{10}^{11}$ (1.3 mmol) in pyridine containing N-methylimidazole (5 equiv.) was added to $\underline{9}$. TLC-analysis, after 20 min at 20°C, showed the formation of dimer $\underline{11}$. Work-up of the crude product, followed by short column chromatography, afforded homogeneous $\underline{11}$ (δ_p 23.69 and 24.34) in a yield of 71% (based on $\underline{8}$). A small amount of dimer $\underline{11}$ was completely deblocked by removing¹² firstly the MPS-group with syn-pyridine-2-carboxaldoxime and N¹,N¹,N³,N³-tetramethylguanidine¹³. The different acid-labile [R¹=2-hexadecyloxy-4',4''-dimethoxytrityl; R²=tetrahydropyranyl and the 2',3'-O-methoxymethylidene (MM)] groups were removed in the following order. Group R¹ was deblocked by a short treatment with aq. HOAc. The acetic acid was removed *in vacuo* and the solution was adjusted to pH 2 (0.1 N HCl: removal of the THP and ring opening of the MM group) and left for 24 h at 20°C. The solution was basified (pH 8) with aq. ammonia and further processed, to give, after purification, homogeneous UpU (δ_p -0.20).

The versatility of agent $\underline{3}$ was further illustrated by the regioselective introduction of a



3'-5'-phosphotriester bond (i.e. $\underline{13}$) and, also, by the synthesis of dimers $\underline{14}$. Thus addition of $\underline{12}^{14}$ (1.3 mmol) to $\underline{9}$, under the same conditions as used for the preparation of $\underline{11}$, gave dimer $\underline{13}$ (δ_p 24.69 and 25.19) in 73% yield. The conversion of $\underline{13}$ into dimers $\underline{14}$ (R³=H and R³=-CH₂CH₂CN) was performed in two different ways. In the first approach, a solution of $\underline{13}$ (1 mmol) in pyridine was phosphorylated with $\underline{3}$ (1.15 mmol) to give, after 20 min at 20°C, intermediate $\underline{14}$ (R³=BT). The latter was hydrolyzed with water to afford $\underline{14}$ (R³=OH), which could be isolated by extraction, followed by precipitation, as the triethylammonium salt in 85% yield. In the second approach, intermediate $\underline{14}$ (R³=BT) was converted *in situ* by the addition of an excess of β -cyanoethanol into $\underline{14}$ (R³=-CH₂CH₂CN), which could be isolated, after column chromatography, in a yield of 81%. Complete deblocking of $\underline{14}$ thus obtained was as follows. Treatment of $\underline{14}$ with N¹,N¹,N³,N³-tetramethylguanidine (9 equiv.) resulted in the instantaneous elimination, as followed from ³¹P-NMR spectroscopy, of the β -cyanoethyl group. Addition of syn-pyridine-2-carboxaldoxime (10 equiv.) to the above mixture released the internucleotidic MPS-protecting group. The MPS-group at the 3'-phosphodiester was now removed by oxidation with iodine, and the R¹ and R² groups by acid. Purification of the crude product afforded UpUp, which was identical, ³¹P-NMR (δ_p 1.30 and -0.18) and TLC-analysis¹⁵ (R_f 0.51), with the product obtained after complete deblocking of $\underline{14}$ (R³=OH). Furthermore, enzymatic digestion of the UpUp compounds, prepared by the two different approaches, were completely digested, as followed by TLC-analysis and monitoring of the enzymatic process by ³¹P-NMR spectroscopy, by RNase to give solely uridine-3'-phosphate (δ_p 1.72; R_f 0.63).

In conclusion, we believe that the easily accessible agent $\underline{3}$ promises to be very convenient for the introduction of polyphosphate functions at the terminal ends of nucleic acids. It also seems to be an alternative for the existing procedure¹² in which the cyclohexylammo-

nium salt of S,S-diphenylphosphorodithioate has been used for the preparation of ArS-protected intermediate internucleotidic phosphotriester linkages. At present, we are studying in detail¹⁶ the scope of the new agent 3 in nucleic acids and sugar chemistry.

ACKNOWLEDGEMENT

This work was supported by the Netherlands Foundation for Chemical Research (SON), with financial aid from the Netherlands Organisation for the Advancement of Pure Research (ZWO).

REFERENCES AND NOTES

1. a) A.G. Hovanessian, R.E. Brown and I.M. Kerr, *Nature (London)*, **268**, 537 (1977); b) K. Miura, *Adv. Biophysics*, **14**, 205 (1981).
2. C.B. Reese and L. Yau, *J. Chem. Soc., Chem. Comm.*, 1050 (1978).
3. G.A. van der Marel, C.A.A. van Boeckel, G. Wille and J.H. van Boom, *Nucl. Acids Res.*, **10**, 2337 (1982).
4. S.S. Jones and C.B. Reese, *J. Amer. Chem. Soc.*, **101**, 7399 (1979).
5. A.F. Cook, M.J. Holman and A.L. Nussbaum, *J. Amer. Chem. Soc.*, **91**, 1522 (1969).
6. a) C. Schattenkerk, G.M. Visser, G.A. van der Marel and J.H. van Boom, *Nucl. Acids Res.*, **11**, 7545 (1983); b) G.M. Visser, C. Schattenkerk and J.H. van Boom, *Recl. Trav. Chim. Pays-Bas*, **103**, 165 (1984); c) G.M. Visser, R. Keemink, C. Schattenkerk, B. Kraal and J.H. van Boom, *Nucleosides and Nucleotides*, **3**, 277-286 (1984).
7. J.G. Moffatt and H.G. Khorana, *J. Amer. Chem. Soc.*, **83**, 649 (1961).
8. a) K. Sasse in *Methoden der Organischen Chemie (Houben-Weyl)*, Band XII/2, *Organische Phosphorverbindungen*, Teil 2, vierte auflage, **11**, pg 597, ed. E. Müller, George Thieme Verlag, Stuttgart, F.R.G. (1964); b) L.N. Shitov and B.M. Gladstein, *Z. Obsc. Khim.*, **38**, 2340 (1968).
9. G.A. van der Marel, Ph.D. Thesis, Leiden University (1982).
10. J.H. van Boom and C.T.J. Wreemann in *Oligonucleotide Synthesis: A Practical Approach*, ed. M.J. Gait, I.R.L. Press, Oxford, U.K. (1984), 153-183.
11. B.E. Griffin, M. Jarman, C.B. Reese and J.E. Sulston, *Tetrahedron*, **26**, 2301 (1967).
12. T. Kamimura, M. Tsuchiya, K. Urakami, K. Koura, M. Sekine, K. Shinozaki, K. Miura and T. Hata, *J. Amer. Chem. Soc.*, **106**, 4552 (1984) and references cited therein.
13. C.B. Reese and L. Zard, *Nucl. Acids Res.*, **9**, 4611 (1981).
14. B.E. Griffin, M. Jarman and C.B. Reese, *Tetrahedron*, **24**, 639 (1968).
15. Eluens: aq. ammonium acetate (1 M): ethanol, 3:7, v/v. Solid phase: DC-Alufolien Cellulose F (Merck).
16. Experimental evidence so far obtained indicated that agent 3 has the same favourable phosphorylating properties as 2-chlorophenyl-0,0-bis[1-benzotriazolyl]phosphate [C.T.J. Wreemann et al., *Nucleic Acids Res.*, **11**, 8389 (1983) and references cited therein] and, further, that the pyridine-2-carboxaldehyde oximate ion-promoted unblocking of the MPS-group from dimer 11 proceeds four times more slowly than the removal, under the same conditions, of the 2-chlorophenyl group from a similarly protected UpU dimer.

(Received in UK 7 December 1984)